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Assessing the population structure OPEN and genetic variability of Kenyan native goats under extensive production system

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Indigenous goats are important to many livelihoods. Despite this, they are subjected to indiscriminate crossbreeding. This afects their genetic variability which is needed to survive in current regime of climate change. The study assessed population structure and genetic diversity of Galla and Small East African goats (SEA) using pedigree information. A total of 7384 animals, 5222 of the Galla and 2162 of the SEA breeds, born between the years 1983 and 2022, were utilized. Individuals with known parents were defned as reference population. From the results, the maximum generation traced for Galla and SEA populations was 14.6 and 14.5, respectively. However, only 6 and 5 generations for Galla and SEA were complete. Pedigree completeness increased with the increasing number of generations. The average generation interval (*GI***) for Galla and SEA was 3.84 ± 0.04 and 4.4 ± 0.13 years. The average increase in the rate of inbreeding per generation for Galla and SEA was 0.04 and 0.05, with the efective ancestors and founders (***fa/fe***) ratio being same (1.00) for both breeds. Fifty percent (50%) of the genetic variability in the populations was contributed by 3 and 1 ancestor for Galla SEA, respectively. The efective population size (***Ne***) was 5.19 and 4.77 for Galla and SEA. Therefore, the current breeding programs should be changed to avoid future genetic bottlenecks in this population. These fndings ofer an opportunity to enhance the current genetic status and management of Kenyan native goats and other regions with similar production systems.**

In developing countries, rural farmers in small-scale management systems ofen produce native breeds that are adapted to their respective areas. The success of these farmers depends on the ability of breeds to produce efectively for them to enhance productivity. Structured selection and conservation programs are among the proposed remedies that can help improve the genetic progress of these native breeds^{1[,2](#page-8-1)}. One crucial goal of these programs is to maintain genetic diversity within a population^{[3](#page-8-2)}. However, this seems not to work, especially with the extensive production system adopted by most goat farmers in the country. This issue is worsened by inadequate information on the application of sustainable genetic improvement programs as well as constraints in financial resources^{[4](#page-8-3)}. Genetic diversity refers to the variety of genotypes or phenotypes available in a given population. Tis includes all the physical, physiological, and behavioural distinctions observed among individuals and populations^{[5](#page-8-4)}. Genetic diversity is essential in matters related to population evolution and adaptation to climate change as well as sustainability within population genetic selection^{[6](#page-8-5),[7](#page-9-0)}. Despite the molecular techniques being employed to assess genetic diversity and relationships within and among herds⁷⁻⁹, pedigree analysis remains the most convenient and cost-efective method. Tis is particularly true for tracking changes in genetic variability, once genomic relationships among founder animals are established 10^{-12} 10^{-12} 10^{-12} .

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Kenya is home to native goat populations widely distributed across diverse agroecological regions¹³. These goats play a vital role in supporting rural household economies by providing food and nutritional security through milk and meat supply, generating income from surplus livestock sales, and serving as insurance against unforeseen risks^{[1](#page-8-0)}. Furthermore, they hold significant cultural values and contribute to climate-smart agricul-tural practices, enhancing farmers' resilience to challenges posed by climate change^{[1](#page-8-0),[14](#page-9-5)}. According to the most recent estimate by[15,](#page-9-6) the number of goats was approximately 32.7 million in 2022 which was higher than that of cattle (23.5 million) and sheep (23.8 million) as shown in Fig. [1](#page-1-0). This figure emphasizes on how these goats are preferred by most households as compared to sheep and cattle.

Among the native goat breeds in Kenya, the Galla goat (Fig. [2A](#page-1-1)), also known as the Somali or Boran goat, is a hardy breed found in the arid and semi-arid regions of the country. These regions experience minimal precipitation, high temperatures, poor-quality feed supplies, and a high prevalence of livestock diseases¹⁶. Galla goats are well-adapted to these harsh environmental conditions due to their long and tall body structure. They are predominantly white and primarily raised for meat production, with some communities utilizing them for milk production^{[17](#page-9-8)}. The Small East African (SEA) goat breed (Fig. [2](#page-1-1)B), indigenous to Eastern and Central Africa, is another small ruminant important genetic resource in Kenya. Although highly prolifc, hardy, and adaptable to the local environment, SEA goats are less proftable for breeders, as they typically reach a market weight of only 20 kg by the age of 2 years^{[18](#page-9-9)}.

Despite their importance, these native breeds have ofen been neglected by breeders due to their lower productivity and slow growth rate. To improve their performance, the breeds are commonly exposed to indiscriminate crossbreeding with international breeds¹³. However, this practice may affect the adaptive features of these native goats thus leading to extinction or the dilution of their adaptive characteristic[s19,](#page-9-10)[20.](#page-9-11) To prevent this from happening and ensure the preservation of their valuable traits, it is crucial to establish a sustainable breeding program that will efectively safeguard and manage these trait[s14](#page-9-5)[,21.](#page-9-12) Tis can only be achieved by understanding the population structure and genetic variability, using the pedigree or molecular information^{[7](#page-9-0)}. Various studies have used molecular data to assess the genetic diversity of these native goat[s13,](#page-9-4)[22–](#page-9-13)[25.](#page-9-14) But so far, no studies have been done to assess the genetic diversity of Galla and SEA goats using pedigree data which is readily available. Tis study intends to fll this knowledge gap by examining the population structure and genetic variability of Galla and SEA goats under extensive production system. By evaluating their genetic structure and diversity using the available pedigree data will help in determining ways to develop an efective breeding strategy that aims at conservation and sustainable management of Galla and SEA goats in Kenya.

Figure 1. Population trend for goats, sheep and cattle in Kenya. *Source:* FAOSTAT, 2022¹⁵.

Figure 2. Kenyan Galla (**A**) and Small East African (**B**) native goat breeds.

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Results

Average number of ofspring produced per buck and does

The total number of bucks and does with offspring were 90 and 3063 for Galla and 50 and 1023 for SEA, respec-tively (Table [1\)](#page-2-0). Among the does that produced offspring only gave rise to an average of 1.61 ± 0.23 kids for Galla and 1.75±0.35 for SEA goats, while the average ofspring produced per buck were 59.95±23.2 and 31.16±15.1 for Galla and SEA, respectively. Even though fewer bucks were used than does, there were more replacement ofspring born to bucks than to does.

Pedigree completeness

Pedigree completeness has a significant impact on the calculation of the inbreeding coefficient (F). For this study, there were 5,222 and 2,162 animals in the relationship matrix for the Galla and SEA breed populations, respectively. Figure [3](#page-2-1) displays the pedigree completeness for both populations. The maximum generation for the Galla and SEA breeds in terms of completeness, was 14.6 and 14.5 respectively. Bucks and does make up the frst parental generation, grand-bucks and grand-does make up the second, and great grand doe and great-grand bucks make up the third generation, and so on. For the Galla and SEA breeds, the percentages of frst, second, third, and fourth-generation pedigree completeness were 98.09%, 85.44%, 69.00%, 47.34%, and 96.99%, 84.86%, 62.9%, 44.3%, respectively. Figure [3](#page-2-1)A,B illustrate how the pedigree completeness increased as the maximum generation increased.

Efective population size (Ne)

The estimates for the mean MG, CG, and EqG for Galla and SEA were 6.9, 2.64, 4.01 and 6.41, 2.91, 4.04, respectively. Teir corresponding inbreeding rates (*ΔF*) were 0.04, 0.13, 0.1% for Galla and 0.05, 0.15, 0.10 for SEA breed, respectively (Table [2](#page-3-0)). The upper and lower limits of Ne for the pedigreed population for Galla and SEA were 3.74, 5.19 and 3.35, 4.77, respectively.

Inbreeding coefficient (F)

For Galla and SEA, the average inbreeding coefficient was 0.75 and 1.07, respectively. SEA recorded a higher F as compared to Galla goats. For both breeds, F increased from second maximum generation (0.19%) to fourteen maximum generation (0.79%) in Galla goats and from second maximum generation (0.21%) to fourteen maximum generation (0.83%) in SEA. For Galla goats, the proportion of inbred animals in each generation increased as the number of generations grew (Table [3](#page-3-1)), while the proportion of inbred animals in SEA reached 100% from the 6th generation and remained constant until the $14th$ $14th$ $14th$ generation (Table 4). The mean AR for both breeds increased with generation (*CG*), with Galla recording 0.04 in generation zero (animals with no parental

Table 1. Number of bucks, does and ofspring born by does and bucks, from the period between 1983 and 2022 for Galla and SEA goats. *NoB* Number of bucks, *BoF* Average number of ofsprings born by bucks per period, *NoD* Number of does, *DoF* Average number of ofsprings born by does per period.

Figure 3. Pedigree completeness for SEA (**A**) and Galla (**B**) goat breeds.

	Galla		SEA	
Type of generation	Λ F	Ne	Λ F	Ne
Complete	0.13	3.74	0.15	3.35
Maximum	0.04	8.56	0.05	8.73
Equivalent	0.10	5.19	0.10	4.77

Table 2. Inbreeding rate (*ΔF*) and efective size (*Ne*) by type of generation for the Galla and SEA breeds.

Table 3. Population parameters for maximum and complete generations for Galla goats.

Table 4. Population parameters for maximum and complete generations for SEA goats. Complete generations (CG) are the number of generations, n, between the individual and the furthest generation when both generations ancestors are known. Maximum generation (MG) is the number of generations separating the progeny from its most distantly known ancestor along each path. Generation (G), Numbers (No), average inbreeding coefficients (F%), percentage of inbred individuals (% inbred), average F for inbred animals (F for inbred), average coefficients of relatedness (AR), and effective size (Ne). Increase in inbreeding (IF), Increase in inbreeding per generation (∆F).

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records) and 0.25 in generation six (Table [3](#page-3-1)), and SEA recording 0.02 in generation zero and 0.82 in generation five (Table [4\)](#page-3-2).

Generation interval (GI)

The generation intervals were obtained from four pathways including buck-son (*Lbs*), buck-daughter (*Lbd*), doe-son (*Lds*), and doe-daughter (*Ldd*) are shown in Table [5](#page-4-0). The mean generation interval (*GI*) from the four pathways obtained was 3.8 and 4.4 years for Galla and SEA and the mean age of parents at the birth of their offspring was 3.94 ± 0.02 years for Galla and 4.56 ± 0.043 years for SEA (Table [5\)](#page-4-0).

The efective number of founders (ƒe) and ancestors (ƒa)

Table [6](#page-4-1) displays the outcomes of the analysis conducted on the probability of the origin of genes. Galla breed had a higher number of founders as compared to SEA breed, 99 versus 61. The effective number of ancestors *(fa)* and the effective number of founders *(fa)* were equal in both breeds, with Galla having $f_e = 7$, and $f_a = 7$, and SEA having f_e = 2, and f_a = 2). This resulted in the same f_a/f_e ratio equal to 1, in both breeds. Number of ancestors explaining 50% of genetic diversity were 3 in Galla breed and 1 in SEA breed.

Figure [4](#page-5-0) shows the cumulative genetic contributions of Galla and SEA goat populations. From the results, both breeds recorded a small number of ancestors which contributed to their genetic pool. In Galla goats, the top three ancestors contributed less than 29.95% of genes. The most important ancestor had 100% contribution. For SEA goats, the least important ancestor made up 62.8% of genes, and the top ancestor had 99.98% impact. Tis graph also shows that each ancestor had a similar impact in both breeds, except for the frst three in Galla goats. Notably, in the Galla goat population, 99 ancestors contributed to 100% of the genetic pool while only 61 ancestors in SEA goats made up 99.98% of genes.

Discussion

Pedigree completeness is an important parameter to be considered in any population. Previous studies have indicated that pedigree completeness significantly impacts the accuracy of inbreeding coefficient (F) calculations and the assessment of inbreeding depression in a population. Tis is because the likelihood of identifying a common ancestor increases with more complete pedigree information²⁶. This is why²⁷ emphasizes the importance of having reliable records of the herd in any given study. In this study, we assessed the population structure and genetic diversity of Kenyan native goats using pedigree information.

Pedigree completeness increased as the number of generations increased. This trend may arise because of lack of parental information in the earlier generations (Generation 0). For this study, animals in generation 0 were considered as founder population.

Table 5. Generation interval and the average age of parents at the birth of ofspring for four diferent paths for Galla goats and SEA.

Table 6. Gene origin for Galla and SEA breeds.

The pattern of pedigree completeness is similar to the estimates reported by²⁸ 52%, 10% and 1.4%, for first, second, and third generation were reported by^{[10](#page-9-2)} in Spanish Murciano-Granadina goats. However, for both studies, the level of pedigree completeness increased as the generation increased. The possible reason for this trend was that identifcation of each animal, pedigree recording, and relationship of each animal were maintained systematically and in an informative way in the nucleus that helped in tracing the pedigree completeness.

The average generation interval (*GI*) was assessed using the four genetic pathways: Buck-son (*Lbs*), Buckdaughter (*Lbd*), Doe-son (*Lds*), and Doe-daughter (*Ldd*). From our study, the SEA goat recorded a higher overall value of generation interval as compared to Galla goats. This could be attributed to the fact that Galla goats tend to have a rapid growth rate as compared to SEA goats which makes them be bred at a younger age. Similar values were reported by^{[29](#page-9-18),[30](#page-9-19)} for South Khorasan Cashmere and Creole Goats in America, however, a higher generation interval of 5.28 years was reported by³¹ in Brazilian Marota goats. Lower figures for the generation interval of 3.3, 2.77, and 3.3 years were reported by^{3[,10](#page-9-2)[,28](#page-9-17)} in Jamunapari, Spanish Murciano-Granadina and Markhoz goats.

In Galla breed, the generation interval was long for Buck progeny pathways as compared to that of Doe progeny pathway (Lss 6.4, Lsd 3.9, Lds 3.9, Ldd 3.7 years, (Table [5](#page-4-0)), while SEA recorded higher generation intervals (over 4 years) in all the four pathways. These results are contrary to those reported by³² in Raeni Cashmere goats and^{[28](#page-9-17)} in Markhoz goats. The longer interval in Buck-progeny pathway reported in Galla goats can be explained using fewer breeding sires for a long period of time without replacement. Longer intervals for the four pathways in SEA compared to Galla (4.4 vs. 3.8 years) could be the result of the use of Galla Bucks and Does at a younger age due to their fast growth rate. Both scenarios can lead to a reduction in genetic diversity.

Shortening the generation interval is one of the methods that can be applied to increase genetic progress and increase the proftability of the farm. In goat breeding, a GI of up to 3 years can help maximize selection response and genetic variation. Therefore, it is recommended that when developing a breeding program, the does should be allowed to mate for the frst time at the average age of 2 years and then, at an interval of 1 year. Bucks should start to be used for breeding at 2 years of age and to be used consecutively for only 2 years^{[33](#page-9-22)}.

The effective number of founders (f e) and ancestors (f a) ratio (f a/ f e) varies depending on the genetic structure of the breed, particularly pedigree depth, breeding methods, and the excessive and repeated use of a particular animal[34.](#page-9-23) Excessive use of only a few animals for breeding can increase the chances of mating between related individuals if no restrictions are applied. The effective number of founders (*fe*) and the contribution of the founders were well-balanced in both breeds, recording similar fgures for efective founders(*ƒe)* and ancestors(*ƒa)* with Galla breed having 7 and 7 and SEA breed 2 and 2, respectively. Despite the (*ƒa/ƒe)* ratio being ideal (1.00) for both breeds, 50% of the genetic variability of the population was determined by a few ancestors. Besides, both the breeds had few ancestors who contributed cumulatively to their overall genetic pool. This normally occurs when the selection of breeding animals is based on physical observation where animals with desirable traits are selected for breeding and retained in the farm for a longer period (selective pressure), resulting in a limited genetic diversity in the population.

Tis scenario may contribute to future genetic bottlenecks if the current mating policy is maintained.

A similar (*ƒa/ƒe)* ratio of was reported by[10](#page-9-2) in Spanish Musiano-Granadina goats. While a higher ratio of 7.10 was reported by[28](#page-9-17) in Iranian Markhoz breed, showing a considerable genetic bottleneck due to the lack of transfer of the alleles to subsequent generations. High or low values of the (*ƒa/ƒe)* ratio always indicate an imbalanced use of Bucks/Does, which poses a risk to the original genetic diversity. When population sufers a demographic decrease, *fe* is overestimated by ignoring some genetic bottleneck effects. Thus³⁵, considered that the estimate of the actual number of ancestors (*ƒa)* may have complementary information to (*ƒe)* as this allows us to consider the loss of genetic variation caused by the unbalanced use of sires, which is adequate to evaluate the loss of genetic diversity 10 .

Besides this, efective number of founders (*ƒe)* were lower than the founder numbers (*ƒ)* in both breeds. Tis may indicate the loss of genetic diversity caused by unbalanced contributions of founders as it was expected that all founders to contribute equally to breeding³⁶. The (fe/f) ratio gives a clear idea about the disequilibrium observed in founders' contribution to the population. Tis ratio was found to be 0.07 for Galla and 0.03 for SEA. Different estimates for this ratio were reported by other authors, such as 0.23 in Cashmere goats³⁷, 0.46 in

Markhoz goats^{[28](#page-9-17)}, and 0.14 in Jamunapari goats^{[3](#page-8-2)}. These findings further explain the fact that although the main objective of a breeding plan was to select superior genes for breeding purposes, it ultimately led to reducing the genetic diversity of the population, as few animals were used in subsequent generations. These findings also explain the increase in the inbreeding coefficient and the higher average relatedness in the population, along with a decline in genetic diversity and allelic loss due to genetic drif. Hence, this population parameter provides useful information in formulating strategies that will minimize inbreeding in a population.

The coefficient of inbreeding (F) measures the number of related alleles that are being produced, the variety of genes that are being lost, and any potential ftness losses linked to a higher risk of phenotypic expression of genetic abnormalities^{[38](#page-9-27)}. As a result, it is advised that the level of inbreeding in any population be kept at a man-ageable level, which is one of the goals of conservation programs^{[39](#page-9-28),[40](#page-9-29)}.

According to 36 , pedigree populations typically have inbreeding rates that are higher than AR rates, because it considers the percentage of a population's whole lineage that descended from a common founder. AR can be used in place of or in addition to inbreeding to predict the long-term inbreeding of a population. As a result, AR can be used to maintain genetic variability in a population by joining animals with low AR values³². As AR reaches zero the genetic diversity in a population is said to be maximized.

For this study, the average F and AR for Galla and SEA breeds were 0.46, 0.18 and 0.44, and 0.47, respectively³ reported a lower figure of F (0.24) in Jamunapari goats in India. This lower value was due to the periodic introduction of external Bucks that were only allowed to be used for 3 years, and strictly served 20 Does per year. Studies on the genetic variability of Iranian Adani goats also reported a lower $F(0.24)^{41}$ $F(0.24)^{41}$ $F(0.24)^{41}$. These low values were attributed to the lack of Bucks registration, as approximately 44.4% of animals had missing Buck records. The average increase in inbreeding per generation for Galla and SEA breeds were 0.04 and 0.05. These rates clearly lie within the 1% increase per generation limit set by FAO in assessing breeds classified as threatened with extinction⁴². However, these low inbreeding values can be attributed to missing and inaccurate pedigree information^{[35](#page-9-24)}. The average inbreeding coefficient increased over generations indicating that the mating program encouraged breeding amongst closely related animals.

The effective population size (Ne) of between 50 and 100 animals or the annual inbreeding rate of 1% should be maintained for the population to be classified as fit⁴². Further, if it is anticipated that the genetic variability within a population will not increase due to mutation, then an efective population size of 500 animals should be maintained^{[5](#page-8-4)}. Reduced effective population size means reduced genetic diversity, and it relates to a variety of negative phenomena, including inbreeding, depression in ftness-related characteristics, and higher fuctuation in selection^{[8,](#page-9-32)32}. The Ne reported in the current study for Galla and SEA populations was way below the two limits (50–100) recommended by FAO. Higher findings were reported by³ in Indian Jamunapari goats and by^{[28](#page-9-17)} in Markhoz goats. However, missing pedigree information can lead to an overestimation of efective population size suggesting that the population is viable and has an adequate genetic variability, mating can be planned to achieve genetic gain without ignoring the relatedness among animals^{[43](#page-9-33)}.

Conclusions

The Kenyan indigenous goat breeds have been neglected and show inadequate genetic diversity, the study revealed a low effective population size, which is below the FAO's recommended guidelines. This indicates a low genetic diversity among the breeds. Additionally, the research revealed that only a few animals made a noteworthy contribution of 50.00% towards the genetic variation within the breed. Hence, the current breeding program may pose a challenge if not changed. Our findings also show a notable rise in inbreeding and relatedness coefficient which may be underestimated due to the incompleteness of pedigree data and the extensive production system. These factors negatively impact the sustainable breeding of the studied goat populations and can also lead to a reduction in genetic variability. Hence, it is imperative to strike a balance between conservation for sustainable animal breeding and attaining a desirable selection response. Maintaining a proper balance is crucial to uphold genetic diversity and avoid inbreeding, securing the survival and adaptability of our native goat breeds. To ensure the diversity and sustainability of these goat breeds, it is important to create a breeding program which is well coordinated by relevant stakeholders in the feld, who can provide useful information to farmers involved in extensive goat breeding, as well as sensitizing them on the importance of conserving the indigenous goats. These fndings ofer an opportunity to enhance the current genetic status and management of native goats in Kenya and other regions with similar production systems. However, the founder population had missing pedigree information, which could be a limitation to this study. Considering this, future studies should involve the use of molecular data, which can provide a more accurate and comprehensive assessment of the genetic variability of these goat breeds.

Material and methods Animal species and data

Data and pedigree information on Galla and SEA were collected from the Kenya stud book and sheep and goat stations. Figure [5](#page-7-0) shows the protocol followed in data selection and processing. A total of 7324 animals (*n*=5224 Galla and $n = 2162$ SEA) born from 1983 to 2022 were analyzed. Information collected included the animal identifcation number (Animal_ ID), Doe number (Mother_ ID), Buck number (Father_ ID), Date of birth, and sex of the animal. The flock was maintained under an extensive management system. Animals were given mineral supplements which were always placed next to their sleeping area and were subjected to average daily grazing hours of 7–8, depending on the weather conditions and routine management. The mating period varied across the year with majority falling between April/May and September/October with corresponding kidding occurring in November/December and April/June. The total number of Bucks and Does with offspring were 90 and 3063 for Galla and 50 and 1023 for SEA, respectively. Among the does that produced ofspring only gave rise to

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Figure 5. Protocol for data selection and processing flowchart.

an average of 1.61±0.23 kids for Galla and 1.75±0.35 for SEA goats, while the average ofspring produced per buck were 59.95 ± 23.2 and 31.16 ± 15.1 for Galla and SEA, respectively.

Statistical analysis

The *ENDOG* software version 4.8^{[44](#page-9-34)} was used to assess various parameters. This software is specifically designed for analyzing populations and genetics using pedigree information. It allows for the assessment of genetic variability and population structure dynamics. The following parameters were analyzed: pedigree completeness, generation interval, inbreeding coefficient, average relatedness, effective population size, effective number of founders and ancestors.

Estimating pedigree completeness

The pedigree completeness indicates how much of the lineage is complete in each generation (the number of complete generations known)^{[45](#page-9-35)}. This analysis was carried out to determine the accuracy of each individual's pedigree information; (a) Number of maximum traced generations (*MG*), defned as the number of generations separating the progeny from its most distant known ancestor along each path. Tose ancestors who did not have parents were regarded as founders and allocated to generation 0, (b) Complete generation equivalent (*EqG*) was calculated as the most distant generation for which all ancestors could be determined, (c) Number of complete generations (*CG*) defned as the number of generations n, between the individual and the furthest generation when both generation ancestors are known and were calculated as:

$$
CG = \left(\frac{1}{2}\right)n
$$

where n is the number of generations separating the individual to each known ancestor ³⁰.

Generation interval (GI)

The generation interval was defined as the average age of parents when their offspring are born^{[38](#page-9-27)}. GI is important because it helps in the identifcation of animals to be used for breeding, as those with shorter GI are more likely to pass on their vital traits quickly to the next generation, thereby contributing to quicker genetic progress 28 28 28 . In this study, the generation interval was assessed using diferent pathways: Buck-son *(Lss*), Buck-daughter (*Lsd*), Doe-son (*Lds*), and Doe-daughter (*Ldd*). The average generation interval of the population was calculated by taking the average of the four pathways using the following formula:

$$
GI = \frac{Lss + Lsd + Ldd + Lds}{4}
$$

Inbreeding coefficient (F) and average relatedness coefficient (AR)

The inbreeding coefficient (F) is the degree of relationship between two individuals who share common ancestors and was evaluated using the formula proposed by[41](#page-9-30). Inbreeding rates per year (*ΔF/y*) and per generation (*ΔF/g*) were derived by regressing F values on year of birth or EqG, respectively,

The average relatedness coefficient (*AR*) evaluates the probability that any randomly chosen offspring in the population originates from a specific animal in the pedigree data⁴⁰. It could be used in place of or in addition to inbreeding to predict the long-term inbreeding of a population. Therefore, AR can be used to maintain genetic variability in a population by joining animals with low AR values²⁸. This coefficient was also computed by employing the methodology described by [45](#page-9-35).

Efective population size (Ne)

Efective population size represents the number of individuals in an ideal population that would result in the same rate of inbreeding as observed in the actual population being studied. It is inversely proportional to the increasing rate of inbreeding^{[46](#page-10-0)}. By regressing individual inbreeding on the mean maximum generation (*MG*), mean complete generation (*CG*), and mean equivalent generation (*EqG*), the efective population size (*Ne*) was calculated. Tis method was preferred as it is used when there is a likelihood of inadequate genealogical data and was done to approximate the lower, higher, and actual bounds of Ne³¹. Individual change in inbreeding Δ Fi was calculated using the formula defined by⁴⁰.

$$
\Delta Fi = 1 - \sqrt[EqGi-1]{1-Fi}
$$

where F_i and E_qG_i are the coefficient of inbreeding and the equivalent complete generation for the individual *I*, respectively. The coefficients of individual increase in inbreeding were averaged, and realized effective population size was estimated as follows.

$$
\mathit{NeF}={\it ^1\!/}_{2\Delta F}
$$

The effective number of founders (fe) and ancestors (fa)

These parameters were analyzed to characterize the genetic variability of the population in terms of gene origin for animals with both parents known (Reference population). Te efective number of founders (ƒe) is the number of animals that, if mated randomly, would produce the same amount of genetic variation as the study population. This parameter considered the animals whose genealogy was known. The calculation was performed by summing up the number of progenies of each founder in the population 34 .

$$
\text{fe} = \frac{1}{\sum_{k=1}^{f} \frac{q2}{k}}
$$

where f was the number of founders and q_k presented the genetic contribution of the k^{th} founder to the reference population 34 .

The genetic contribution of ancestors (f) of the reference population (founders or not) and their expected marginal contribution to the gene pool were assessed using the method proposed by³⁴. This method was estimated using the formula that calculates the likelihood of an ofspring inheriting an allele from a specifc ancestor proposed by³⁰. *ENDOG* software was used for this calculation. while the effective number of ancestors (fa), which is the minimum number of ancestors, not necessarily founders, explaining the complete genetic diversity of the current population was also computed basing on marginal contribution of each ancestor as follows:

$$
fa = \frac{1}{\sum_{j=1}^{a} \frac{q^2}{j}}
$$

where qj is the marginal contribution of an ancestor j, i.e., the genetic contribution made by an ancestor that is not explained by other ancestors chosen previously 30

Ethical declarations

There was no ethical approval needed as the data used for the study was from the records kept by the herdbook.

Data availability

The raw data analysed in this study is the property of the Kenya herd book and sheep and goats project hence are available upon reasonable request through the corresponding author.

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Competing interests

The authors declare no competing interests.

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